PR10_Evolucijska in komparativna genomika ter filogenomika

The DNA Record is a Living Chronicle of Evolution



- Information in genomes is vital to reconstructing the processes and patterns of evolution
- Knowledge of evolution is a powerful guide to interpreting genomes

Understanding Sequences Requires Tools and Evolution



What is a Genome Like?

ACAACCCCTCCACCTCATGTACCTGCGGACTCTCCCAGTCACGGCAGTCCACGTCTGCAACCCCCTAAACCCTCAAAACCGGTT GACGTTCTGTTAGACGAACAACTATGATATATCGACCCCGCCTAAGAACGGAGCCTCTGTCAGTGCTCCAGCTGAACGTAGGCCGCGGG TATGTCCGGAAAAAGATGGGCATTCGGGCCTCTCAGCTCCGCCCTCAGCCAATAGATCAAGATGTTCTCTCAGACCTTCTTCTACTACAG ATCCTCTCCCGGCTCTGGACAATCTGCATTGATAATCAACATCTATAATGCTCCAATCGGCTCAATCAGGTCAGGTGAGGCTGCAAAAGCG CTTACACTCCTGCCTGACTCCTACTTTTCCCAGCCTACCGTGCTTGCCGGCGACTTCAACCTACTACATAGCAGGTGGCAGCCATCACTG CATTGCAGCCCTACCACCTTTGCTGAGCCATTTGTTGACTGGCTTGATCGCCTAGGGCTGGTTCTTATCTCCGAGATAGACCAGCCTACAC ACGATAGAGGCAACGTTCTTGACCTCACTTTCGCCTCCAGCTCCCTAGCACTGGCAGGGTCGAGTACCAGGATAGCAAGTCATTTAGAGT CAACATCAGATCATCGGCCACTCCTCACCACCATGCCATGGAGCCAGAGATTCACAGAGGCAGCTCAGAAACTGAGATTTGATACATTA GACCACCCTCGCTTCCTCTCACTACTCAGTTCCCACCTTGCTGTCATTGAATGCTCAGCTACAACAGAAGAGGGCCTGGACAGTCTAGCT CATGGGTTAACCTTAGCAACTGCTAGTGCGTATAAAGGCTCTGCTAGGAGCTCCTTGGCGCAGGGAATAGGTCAGCCATGGTGGAATATT GACTGCAGAAAAGCGTTGCAAGACTTCCGCTTAGGTCTCTGTTCAAGAAACGACTTCCGTCGGATAACTAGACGGTCTAAATAGCAGTTC TGGCGAGATAAACTTACCGCAGTGACACAGATCAAAGATGTCTTTGACATAAGCAAGTGACATAAGTTTACAGGATCTTATCGAAACCCT CCACTAAACGACCCTTTAAGGCCAAACAGCCCTCCAGCAGGGGCTCTGAATGAGAAACAAGACGTATTAGTCCGTAATCTTCTTCAGAAT ACTGCTGAAGCGGGTGATATTGTCATAGGCTATGGCCTGGGCTGTGGTTGTCAGCCATGCCCTCAACCATAGAACATTCTAGAAGAACCA **GCGATATTCGTTAATACATTATACAGGATTGCCAGTTGAAAACAATACTGCCTACGCCCGTCACAGGTACTGCAGTTTCCAACAAGAATC** AACGCTCGACCCGGCAATTATGGCTCAAGGTTAGACTACGTCCTGTGTAGCCTTGATATGCAAGATTAGTTCTGCGATTTGAATATCTAAG ATTGTCCGGTTTATGCTGTCTTCAAAAATGTTATACGCCTCGGCGAAGAAGAGGTCAACATTAAATGAGCCCTCCTGGGATGTTTAAAGAT **GGCGAGCGTCAGCAGGAATACTCTACTAAATATCTTCTGCCTACATCAGGGCGCTTAATACCAGAATTTAACAAGCGGAGGAGGATCAA GGACATGTTCTTGCGTAAACCATCAGCCAACGTATAGAGACCGACGACGACATCCTGACATTGAGATATTTTACCTCTAGTCAGGAAAA** GGGAACAGCACCCGCTATTTTGGAGAGTGCTGCCAGCGTCATAGCTACCTGCCAGCCTGTAGTAGCTGCTGACAGCACTCAAATGAAAG AAGTTATTCGTAAGAGCTCTCAGAAATATGAGACAGGTTCCCCTGTCTCAGTCCAGTATTTGACATCGGGTTCAGCCCAATCATCAACAC ACAGTGTCTACTATGCCAACGAAACAACCAGCCGCATCCCCTACAAAATCTACCCCAGTTACAGAACCTCCTGCACTGGAAGCATTACTG ACAGCTCCCGCTGGTGAAGCTTCTCCAGGAGAACAGCCAAATTCCGCGACTCCTACAGCTCCCGCTTCACCCCAAAGCAATGATACTATT ATCGATCCCATTGTCAGCAAGGAAGATTGGTCAAAGCTCTTCACTAAAAAGCCCATTCCCAAGTGCGAGGGCCACCAGGAACCATGTTT CAGTCTGACAACTAAGAAGCCTGGCATCAACTGCGGAAGATCGTTCTGGATCTGTTTGAGACCCCTTGGGCCCAGCGGAAACAAGGAAA TGCGATAGTGTAACGATACCCGGTTTTATACTTAGAAGGCTACGAATGGTATGATGATGTATCATGGTTTCAATGATAAGACATTTCGTCAAGT

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ACAACCCCTCCACCTCATGTACCTGCGGACTCTCCCAGTCACAGCTCAGGCAGTCCACTTTGCAACCCCCTAAACCCGGTTT GACGTTCTGTTAGACGAACAACTATGATATATCGACCCCGCCTAAGAACGGAGCCTCTGTCAGTGCTCCAGCTGAACGTAGGCCGCGGG TATGTCCGGAAAAAGATGGGCATTCGGGCCTCTCAGCTCCGCCCTCAGCCAATAGATCAAGATGTTCTCTCAGACCTTCTTCTACTACAG ATCCTCTCCCGCTCTGGACAATCTGCATTGATAATCAACATCTATAATGCTCCAATCGGCTCAATCAGGTCAGGTGAGGCTGCAAAAGCG CTTACACTCCTGCCTGACTCCTACTTTTCCCAGCCTACCGTGCTTGCCGGCGACTTCAACCTACTACATAGCAGGTGGCAGCCATCACTG CATTGCAGCCCTACCACCTTTGCTGAGCCATTTGTTGACTGGCTTGATCGCCTAGGGCTGGTTCTTATCTCCGAGATAGACCAGCCTACAC ACGATAGAGGCAACGTTCTTGACCTCACTTTCGCCTCCAGCTCCCTAGCACTGGCAGGGTCGAGTACCAGGATAGCAAGTCATTTAGAGT CAACATCAGATCATCGGCCACTCCTCACCACCATGCCATGGAGCCAGAGATTCACAGAGGCCAGCTCAGAAACTGAGATTTGATACATTA GACCACCCTCGCTTCCTCACTACTCAGTTCCCCACCTTGCTGTCATTGAATGCTCAGCTACAACAGAAGAGGGCCTGGACAGTCTAGCT CATGGGTTAACCTTAGCAACTGCTAGTGCGTATAAAGGCTCTGCTAGGAGCTCCTTGGCGCAGGGAATAGGTCAGCCATGGTGGAATATT GACTGCAGAAAAGCGTTGCAAGACTTCCGCTTAGGTCTCTGTTCAAGAAACGACTTCCGTCGGATAACTAGACGGTCTAAATAGCAGTTC TGGCGAGATAAACTTACCGCAGTGACACAGATCAAAGATGTCTTTGACATAAGCAAGTGACATAAGTTTACAGGATCTTATCGAAACCCT CCACTAAACGACCCTTTAAGGCCAAACAGCCCTCCAGCAGGGGCTCTGAATGAGAAACAAGACGTATTAGTCCGTAATCTTCTTCAGAAT ACTGCTGAAGCGGGTGATATTGTCATAGGCTATGGCCTGGGCTGTGGTTGTCAGCCATGCCCTCAACCATAGAACATTCTAGAAGAACCA GCGATATTCGTTAATACATTATACAGGATTGCCAGTTGAAAACAATACTGCCTACGCCCGTCACAGGTACTGCAGTTTCCAACAAGAATC AACGCTCGACCCGGCAATTATGGCTCAAGGTTAGACTACGTCCTGTGTAGCCTTGATATGCAAGATTAGTTCTGCGATTTGAATATCTAAG ATTGTCCGGTTTATGCTGTCTTCAAAAATGTTATACGCCTCGGCGAAGAAGAGGTCAACATTAAATGAGCCCTCCTGGGATGTTTAAAGAT GGCGAGCGTCAGCAGGAATACTCTACTAAATATCTTCTGCCTACATCAGGGCGCTTAATACCAGAATTTAACAAGCGGAGGAGGATCAA **GGACATGTTCTTGCGTAAACCATCAGCCAACGTATAGAGACCGACGACGACGACATCCTGACATTGAGATATTTTACCTCTAGTCAGGAAAA** GGGAACAGCACCCGCTATTTTGGAGAGTGCTGCCAGCGTCATAGCTACCTGCCAGCCTGTAGTAGCTGCTGACAGCACTCAAATGAAAG AAGTTATTCGTAAGAGCTCTCAGAAATATGAGACAGGTTCCCCTGTCTCAGTCCAGTATTTGACATCGGGTTCAGCCCAATCATCAACAC ACAGTGTCTACTATGCCAACGAAACAACCAGCCGCATCCCCTACAAAATCTACCCCAGTTACAGAACCTCCTGCACTGGAAGCATTACTG ACAGCTCCCGCTGGTGAAGCTTCTCCAGGAGAACAGCCAAATTCCGCGACTCCTACAGCTCCCGCTTCACCCCCAAAGCAATGATACTATT ATCGATCCCATTGTCAGCAAGGAAGATTGGTCAAAGCTCTTCACTAAAAAGCCCCATTCCCAAGTGCGAGGGCCACCAGGAACCATGTTT CAGTCTGACAACTAAGAAGCCTGGCATCAACTGCGGAAGATCGTTCTGGATCTGTTTGAGACCCCCTTGGGCCCCAGCGGAAACAAGGAAA TGCGATAGTGTAACGATACCCGGTTTTATACTTAGAAGGCTACGAATGGTATGATGTATCATGGTTTCAATGATAAGACATTTCGTCAAGT



Transposor

Protein Binding Site

Exon Intron



Evolucijska oz. komparativna genomika (**Evolutionary and Comparative genomics**)

Evolutionary Genomics

Just as molecular evolution is at the scientific core of molecular biology and genetics, evolutionary genomics is at the core of genomics

Where did genes and genomes come from? (How and why?) A sequence by itself is nearly useless What restrictions are placed on sequences as they change over time? What potential do they have for non-functional change? What potential do they have for malfunction, and can we predict it? How were new molecular innovations created? How do residues interact to effect functional stasis or change? How did diverse molecules work together to create physiological change? What did ancestral molecules look like and how did they function?





Comparisons of Genomes atDifferentPhylogeneticDistances Are Appropriate toAddress Different Questions



Ortholog identification a prerequisite to genomic studies

Current methods for ortholog finding

Pair-wise sequence comparison

- Best bi-directional BLAST hits
- Focuses on one-to-one orthologs (no duplications)

Hit clustering methods

- Detect clusters in graph of pair-wise hits
- Difficulty to separate large connected components

Synteny methods

- Detect conserved regions, stretches of nearby hits
- Genome alignment methods focus on best hits

Phylogenetic methods

- Phylogeny of family clusters orthologs near each other
- Traditionally applied to specific families (not genome-wide)



What is comparative genomics

There are many ways that genomes can be compared

- Whole genome
 - Genome size
 - Genome alignments
 - Synteny (gene order conservation)
 - Gene number
 - Anomalous regions
- Gene-centric
 - Gene families and unique genes
 - Gene clustering by function
- Gene sequence variations
 - Codon usage, SNPs, inDels, pseudogenes

Why Comparative Genomics?

- 1. Conservation over long evolutionary distances suggests functional constraints
- 2. Lack of conservation over short distances may be indicative of adaptive evolution
- 3. Helps us identify both coding and non-coding genes and regulatory elements
- 4. Characterizing the differences between organisms reveals mechanisms of change
- 5. Allows us to achieve a greater understanding of vertebrate evolution
- 6. Leveraging knowledge between species for annotation and inference of function
- 7. Tells us what is common and what is unique between different species at the genome level
- 8. The function of human genes and other regions may be revealed by studying their counterparts in simpler model organisms

Comparative genomics

 Discover what lies hidden in genomic sequence by comparing sequence information.

Main areas

- □ Whole genome alignment
- □ Gene prediction
- □ Regulatory element prediction
- □ Phylogenomics
- □ Pharmacogenetics
- Affected by evolutionary aspects
 - Mutational forces (introduce random mutations)
 - Selection pressures
 - \Rightarrow Ratio of non-synonymous to synonymous substitutions
 - \Rightarrow Mutation rates lower or higher than neutral



Comparing sequences, methods.

- Pairwise comparison of sequences (alignments)
 - □ proteins or genes
 - variety of local alignment tools like BLAST, Smith-Waterman etc.
 - multiple sequence comparisons (ClustalW, Muscle etc.)
 - results may be dependent on alignment settings

<u>>HS.239752</u> #DE UNIGENE #DE GNL UG HS_S1731711 HOMO SAPIENS NUCLEAR RECEPTOR SUBFAMILY 2, GROUP F, MEMBER 6 (NR2F6), MRNA /CDS=(132,1346) /GB=NM_005234 /GI=20070198 /UG=HS.239752 /LEM=L830 #OS HOMO SAPIENS (HUMAN) #IT UNIGENE Length = 1830	L
Score = 206 bits (104), Expect = 1e-51 Identities = 334/408 (81%), Gaps = 2/408 (0%) Strand = Plus / Plus	
Query: 626 tgggcatcgagaacatctgcgagctggccgcgcctgctttcagcgccgtcgagtggg 685 	
Query: 686 cccgcaacatccccttcttcccggatctgcagatcaccgaccaggtgtccctgctacgcc 745 	
Query: 746 tcacctggagcgagctgttcgtgctcaacgcggcccagtgctctatgccgctgcacgtgg 805 	

Sequence: <u>REFS</u> Sequence: <u>REFS</u>	BEQ:NM_012387 BQ:NM_011059 BEQ:NM_017230 2) multiple sequence alignment
NM_012387 NM 011059	AGAAGGCACGCTCCTACCCGGCAGCCGGGTGTCCCCCAAGGAGATCTGGGGAGCCGGGGAG
NM_017230	GAG
NM 012387	AGCCAGAGGGACGAGC-TAGCCCGACGATGGCCC-
NM 011059	CAGCTGGGGCTTGTGGCTGCTGGGCACCCAAGCTGACCCGACCAGGAGACTGGAAGCTCG
NM_017230	T-GCTGGGTTTGGTGGCCAAAGCCCCCATCCTAGGCCGGC-ATGTCTCT ** * * * * * *** * ** * * * *
NM_012387	AGGGGACATTGATCCGTGTGACCCCAGAGCAGCCCACCCATGCCGT
NM_011059	ACACAGGATGGCCTCCCCGAGAGCTGTGCAGCTGTCTCTGAGAAAGCCGACTCACGCTGT
NM_017230	GCAGAGGACTG-TACGTGTATCCCTGGAACACCCAACCAGCGCTGT * *** * * * * * * * * * * * * * * ** **
NM_012387	GTGTGTGCTGGGCACCTTGACTCAGCTTG-ACATCTGCAGCTCTGCCCCTGAGGAC-TGC
NM_011059	GTGCGTGGTAGGTGTGGAGACACTG-GTGAACGTCTATAGTGATGTGCCCAAAGGTGCCA
NM_017230	GTGTGTGGCAGGGGTTGAGAC-CATCGTGGACATTTATGGGTCGGTTCCTGAGGGCACAG
—	*** *** ** *** * * * * * * *

Comparing sequences, methods.

Whole genome comparisons

- □ Large stretches of sequence
- Divergence up to 450Mya (*fugu*-human) with sufficient similarity remaining.
- □ BLAT, BLASTZ, Phusion/BlastN
 - Seeding strategy \rightarrow alignment extension \rightarrow gapped alignments





Gene prediction

- Comparing sequences has contributed enormously to the accuracy of gene prediction.
- Evidence based method.
 - □ Use cDNAs, ESTs and proteins from various organisms.
 - □ Apply gene feature rules.



Gene prediction

De novo methods.

- □ Alignment of genomic sequences
- □ Splicing rules and other gene features

De novo gene prediction by comparing sequences attempts to model a negative selection of mutations. Areas with less mutations are conserved because the mutations where detrimental for the organism.

Prediction of similar proteins in both genomes.



Regulatory element prediction

- The complexity of higher eukaryotes and their relatively low number of genes can be explained partially through the importance of transcriptional regulation.
- Identification of RE's will have an extensive impact in understanding gene expression patterns (expression intensity, tissue specificity), relations within expression patterns and inferring biological systems or networks.

Regulatory element prediction

- No formal models for regulatory motifs
- Attempt to find conserved regions or motifs based on the global alignment of similar sequences of different organisms (*phylogenetic footprinting*).
 - □ Which species to compare? Evolutionary distance?
 - □ What regions around gene models to investigate? 5' and 3' flanking regions, introns?
 - □ Take expression patterns into account?
 - $\hfill\square$ How does evolution affect RE's?



Position on chr 19 (Mb)

TRENDS in Ecology & Evolution

Identifying DNA conserved sequence elements. Alignment of genome sequences and tools for visualizing those alignments have proven remarkably useful in identifying how evolution has resulted in the conservation of genetic elements and the erasure of nonessential sequence. The figure shows a multi-species alignment of a homologous region to chr 19 in humans spanning the gene encoding apolipoprotein E.

Extreme conservation in enhancers that are shared by human and fish



Comparative genomics at the vertebrate extremes

- Intraspecies sequence comparisons allow identification of species specific sequences
 - Phylogenetic shadowing
 - Requires high rate of polymorphism

- Comparison among primates show human specific sequences
 - Analysis of regulatory sequence of ApoA (involved in human heart disease)



A. Mutation rate analysis of *Ciona intestinalis* 5` region of the *forkhead* gene. B. Validation of identified potential regulatory elements in *Ciona* larvae.

Finding functional regions through genome sequence comparisons

- "Phylogenetic footprinting"
 - Comparing sequences of divergent genomes to find functional regions, eg. Human-mouse
 - Will not reveal elements if background divergence is low
 - Thus cannot reveal novel functional elements in recent lineages, such as great apes
- "Phylogenetic shadowing" (D. Boffelli et al. Science 2003)
 - Using comparisons of close relatives to find regions of conservation
 - Allows detection of elements with varying levels of constraint
 - Eg. Comparing humans to other apes and monkeys





Phylogenetic shadowing to find conserved regions



Whole Genome Alignments

- Functional sequences often evolve more slowly than non-functional sequences, therefore sequences that remain conserved *may* perform a biological function.
- Comparing genomic sequences from species at different evolutionary distances allows us to identify:
 - Coding genes
 - Non-coding genes
 - Non-coding regulatory sequences







Analysis of Comparative Sequence Data

Sequence Identification of Functional Elements

- Coding Sequences (i.e. Genes)
 - Relatively EASY to identify
 - Basic understanding of the 'language'
 - Complementary datasets available (ESTs, cDNAs)
- Non-Coding Functional Sequences
 HARD to identify
 - Very little idea of what to look for
 - Virtually no complementary datasets



 Find sequences that have diverged less than we expect

These sequences are likely to have a functional role

 Our expectation is related to the time since the last common ancestor





Comparative Sequence Tracks



Whole genome comparison

Conservation of syntemy!

□ Cross-reference of any genetic traits (diseases!) from one organism (eg mouse) to genes in the syntenic regions in the other organism (eg human).

Genome expansion and contraction

Genome duplications, segmental duplications: important mechanism for generating new genes.

(G+C) content, CpG islands

□ Reflect different mutational or DNA repair processes?

Repeats

- □ Transposable elements are a main force in reshaping genomes. TE's (or remainders thereof) can be used to measure evolutionary forces acting on the genome.
- □ Neutral mutation rate.

Extensive conservation of gene order in multiple species



Comparing Genome Size The 'C-value paradox'



Genome size does NOT correlate with organismal complexity

Why Are Some Genomes So Large?

- There is no clear correlation between genome size and genetic complexity.
- **C-value** The total amount of DNA in the genome (per haploid set of chromosomes)
- C-value paradox The lack of relationship between the DNA content (C-value) of an organism and its coding potential.







Genome sizes and C-values

Archaea:



The forces affecting genome size evolution (Petrov, TIG 2001)







Processes that generate qualitative and quantitative variation in gene number and overall DNA content in plant nuclear genomes. Small arrows indicate minor events, and large arrows indicate large events.

Current Opinion in Genetics & Development





- Finding information from whole-genome sequencing projects
 - DNA sequence reads

low

high

Information value

- Assembled genomic DNA sequences
- Annotated genes (RNA genes + proteinencoding genes)
- Repeats, transposable elements
- Integrated platform providing both sequence data and functional genomics data

What can we learn from crossspecies comparisons?

Genome conservation

 transfer knowledge gained from model organisms to non-model organisms

Genome variation

 understand how genomes change over time in order to identify evolutionary processes and constraints

Detection of functional elements

- Coding elements (e.g. exons)
- Conserved non-coding sequences / elements
Človeške bolezni in komparativna genomika



Nature Reviews | Genetics

Uporaba komparativne oz. evolucijske genomike



A multidisciplinary perspective on language evolution

The genomic structure of human **forkhead box P2 (FOXP2)**, showing the location of mutations that cause verbal dyspraxia, which are distinct from sites of evolutionary substitution in the human lineage (filled rectangles, coding exons; white rectangles, non-coding exons). The red bar indicates genomic regions that show evidence of a selective sweep.



Evolutionary genomics of SRGAP2 and its human-specific paralogs.

(A) Evolutionary genomic analysis performed by the Eichler's group (Dennis et al. 2012) revealed that SRGAP2 has undergone three human-specific duplications. The first one (SRGAP2B) occurred approx. 3.4 million years ago (mya) and is a partial duplication including only the first nine exons (total of 22 coding exons in the ancestral copy SRGAP2A). The duplication containing large segmental SRGAP2B duplicated approx. 2.4 mya to generate SRGAP2C which also contains the first nine exons. A more recent duplication (SRGAP2D) containing only the first two exons emerged approx. 1 mya. Based on expression analysis as well as copy number variation in the general human population reveals that SRGAP2B and SRGAP2D are most likely pseudogenes. On the other hand, SRGAP2C encodes for a truncated protein (partial F-BAR domain) highly expressed in neurons during human brain development and as a strong antagonist of ancestral acts SRGAP2A, leading to neoteny during spine maturation and increased spine density in vivo.

A distal enhancer and an ultraconserved exon are derived from a novel retroposon



DRG UN TORG

Conserved sequences and the evolution of gene regulatory signals

Potential roles of CNEs in gene regulation.

Conserved non-coding sequence elements (CNEs) might have a diverse array of functions related to gene regulation and maintenance of chromosome structure. Some roles for CNEs include (a) acting as cis-regulatory elements; for example, as an enhancer, silencer or insulator to modulate transcription of an adjacent gene; (b) interacting with components of chromatin remodelling complexes, which create 'open' structures that promote active transcription or 'closed' structures that inhibit it; and acting in concert to bridge regulatory elements located at long distances, either (c) on the same chromosome or (d) on another chromosome.

Identifying functional sequences by comparative genomics approaches

The impact of evolution is most readily seen in the pattern of sequence conservation and variation among species. Sequencing of the mouse and rat genomes has revealed remarkable features of evolutionary conservation. About 5% of each genome is under selective constraint, of which only about 1.5% is protein-coding. This conservation extends throughout mammalian species. The importance of gene regulation is highlighted by the fact that two-thirds of the sequence conserved among mammals is not protein-coding. These 'conserved non-coding sequence elements' (CNEs) are largely unique in each genome (i.e. non-repetitive), and, to date, no readily recognizable clusters, classes or subdivisions have been defined that might be useful in further characterizing them.



Mapping of conserved RNA secondary structures predicts thousands of functional noncoding RNAs in the human genome



GTCAGAATAATGTCAAAGTC

Lineage-specific expansions: Transcription Factor Families Have Much Higher Expansion Rates in Plants than in Animals

After examining the lineage-specific expansion of TF families in two plants, eight animals, and two fungi, we found that TF families shared among these organisms have undergone much more dramatic expansion in plants than in other eukaryotes. The high rate of expansion among plant TF genes and their propensity for parallel expansion suggest frequent adaptive responses to selection pressure common among higher plants.

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Species		MY	DN 11.	A-bin 242 21-0	3HA CB	PAT	PHI PHI	HMF HOP	neob HSF	ox on DN	A.bir	iD _G RÍ	TF. GA	(ALL)	`cB ⁵	, t.8	pot Élt	TDP Tub	'ct	^T AP	L NR	K SBF	FEIN	^{و رو}		5 A.S	DO ^t A	3BY EIN	30U	ist ³ CRI	5 FL	LF NA	4 ANTC
Saccharomyces	erevisiae	i —	46		4	6	14	7	5	8	2	4	10	7	2																		
Candida albicans		16	52	27	7	2	15	3	7	7	2	2	9	11	3																		
Schizosaccharon	iyces	13	22	35	6	5	17	2	2	5	2	3	5	4	2																		
Neurospora crass	a	16	75	35	5	16	14	6	2	9	3	2	6	14	2																		
Fusarium gramin	earum	12	95	30	5	20	15	10	3	10	2	2	6	16	2																		
Magnaporthe gris	ea	12	71	29	5	18	11	4	3	10	з	1	7	9	2																		
Cryptococcus ne	oformans	7	37	28	6	6	16	5	4	12	1	2	7	9	2																		
Ustilago maydis		11	33	28	7	12	13	6	4	5	2	2	8	12	2																		
Encephalitozoon	cuniculi	9	11	15	5	1	3	10	2	1		1	2			1	2	1															
Homo sapiens		31	641	195	8	17	59	204	5	29	13	5	12	90	2	65	8	4	1					2									_
Gallus gallus		29	279	133	6	20	47	126	4	21	10	2	7	70	1	14	8	3	2					1									
Takifugu rubripes	;	27	362	181	7	17	74	214	2	38	15	8	13	102	2	59	10	5	1					2									
Ciona intestinalis		18	166	69	6	7	36	78		1 1	6	2	2	39	1	9	5	1	1					1									
Anopheles gamb	ae	11	322	56	15	6	55	80	1	7	5	2	4	44	1	6	2	1	1					1									
Drosophila melar	ogaster	17	289	70	6	20	35	93	1	9	6	2	6	49	2	6	3	2	2														
Caenorhabditis b	riggsae	13	117	46	14	9	17	64	1	7	3	2	5	21	1	6	3	2	1														
Caenorhabditis e	egans	15	168	90	6	13	24	83	1	9	4	2	12	42	2	15	4	2	1														
Dictyostelium dis	coideum	28	27	48	4	6	8	9		15	1	3	11			27	1				1												
Entamoeba hydro	lytica	36	21	38	4		1	2	8	2		2	1		1	2	L					L					L						
Arabidopsis thali	ana	280	137	373	27	29	63	87	24	73	9	107	29	147	11	29	8	11	8	145	73	15	14	6	83	36	4	6	19	32	1	101	2
Oryza sativa		286	154	340	31	28	58	96	32	95	7	74	32	174	15	34	9	15	12	167	104	16	9	6	91	34	7	11	7	56	1	133	2
Chlamydomonas		22	10	37	4	13	32	3	2	8	2	1	6	8		1	2			15	1	11	6										
Thalassiosira pse	udonana	35	2	53	6	3	22	5	24	5							3	3	7	9													
Plasmodium falc	parum	7	8	22	2	1	5				1					3		1															
 Trypanosoma bru 	cei	1	28	18	1											2																	
Giardia lamblia		11	19	33	2		3				1					1	1				1												

Evolutionary dynamics of vertebrate olfactory receptor (OR) genes



Estimating the tempo and mode of gene family evolution from comparative genomic data

Evolutionary Genomics of Nuclear Receptors: From Twenty-Five Ancestral Genes to Derived Endocrine Systems





Filogenomika/Phylogenomics

Phylogenomic analysis

- Major step in PA is to infer four main events in the evolutionary history of gene family:
 - 1. Gene origin
 - 2. Gene duplication
 - 3. Horizontal gene transfer (HGT)
 - 4. Gene loss
- Evolutionary distribution pattern (EDP)
 - determined by overlaying gene presence/absence information onto an evolutionary tree of species.
 - EDP reveal a great deal about the evolutionary history of particular genes
- Uneven distribution patterns
 - are difficult (scattered presence and absence throughout the tree)
 - HGT or gene loss.
- Gene is present in only one subsection of the species tree:
 - originated in that subsection.

Phylogenomics

(Eisen, Genome Res. 1998)





- A combination of :
 - genomics (study of function and structure of genes and genomes)
 - molecular phylogenetics (study of evolutionary relationships among organisms)
- Two different aspects :
 - using phylogenetic data to infer functions for DNA and protein sequences
 (Eisen. Phylogenomics: improving functional predictions for uncharacterized genes by evolutionary analysis. Genome Res. 1998)
 - using genomic data to infer phylogenetic relationships (species trees) and to gain insights into the mechanisms of molecular evolution (O'Brien and Stanyon, Phylogenomics, Ancestral primate viewed, Nature 1999)

Phylogenomics: three main axes



Evolutionary history of genomes and biological systems

Phylogenomics

The use of phylogenetic principles to make use of genomic data



Methods of phylogenomic inference

The flowchart shows steps in the inference of evolutionary trees from genomic data. Genomic information is obtained by large-scale DNA sequencing. In general, sets of orthologous genes are then assembled from specific sets of species for phylogenetic analysis. This homology or orthology assessment is a crucial step that is almost always based on simple similarity comparisons (for example, BLAST Most methods for searches). used the subsequent reconstruction of phylogenetic trees are either sequence-based or are based on whole-genome features.



Phylogenomics and the tree of life

A schematic representation showing recent advances and future challenges of the phylogenomic approach for resolving the main branches of the tree of life. This tree aims to represent a consensus view on evolutionary relationships within the three domains — Bacteria, Archaea, and Eukaryota — with hypothetical relationships indicated as dashed lines. The main branches that have been identified (purple) or confirmed (yellow) by phylogenomics are indicated. Blue dashed lines underline putative phylogenetic hypotheses that have been indicated by phylogenomic studies and need further investigation. The main uncertainties for which the phylogenomic approach might provide future answers are pinpointed by red circles. Note that most of the progress brought about by the phylogenomic approach has been realized at a smaller taxonomic scale for land plants, and for placental mammals within the metazoans (see main text). The two well-recognized endosymbiotic events involving bacteria that gave rise to eukaryotic organelles (mitochondria and chloroplasts) are indicated by arrows (blue and green, respectively). Note however, that other horizontal gene transfers and gene duplication events are not represented in this organismal tree, although they do constitute important aspects of genome evolution.

Phylogenomics and the evolutionary history of biological systems

